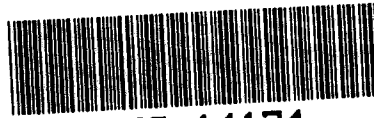




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Elf Atochem North America, Inc.
2000 Market Street
Philadelphia, PA 19103-3222
Tel.: 215.419.7000

8EHQ - 0598 - 14174



8EHQ-98-14174

May 6, 1998

UPS NEXT DAY DELIVERY

Document Control Office (7407)
Office of Pollution Prevention and Toxics
U.S. Environmental Protection Agency
401 M St., S.W.
Washington, D.C. 20460
Attn: Section 8(e) Coordinator

Subject: TSCA Section 8(e) Submission



88980000141

Dear Sir/Madam:

Elf Atochem North America, Inc. (Elf Atochem) is submitting two primary eye irritation studies in rabbits to the Environmental Protection Agency (EPA) pursuant to Toxic Substances Control Act (TSCA) Section 8(e). One study provides information on sodium hexafluoroantimonate (CAS Registry Number 16925-25-0), while the other study provides information on lithium tetrafluoroborate (CAS Registry Number 14283-07-9). These studies do not involve effects in humans.

Nothing in this letter or the enclosed study reports is considered confidential business information of Elf Atochem.

The results of these studies showed both test materials to be corrosive to rabbit eyes.

Further questions regarding this submission may be directed to me at (215) 419-5890.

Best Regards,

Debra Randall

Debra Randall, DABT
Product Safety Manager

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98 MAY 12 PM 12:35

RECEIVED
OPPT NCIC

Contains No CBI

**A PRIMARY EYE IRRITATION STUDY IN RABBITS
WITH SODIUM HEXAFLUOROANTIMONATE**

FINAL REPORT

Author

Deborah A. Douds, M.S.

Study Completed on

December 23, 1997

Performing Laboratory

Springborn Laboratories, Inc. (SLI)
Health and Environmental Sciences
640 North Elizabeth Street
Spencerville, Ohio 45887

SLI Study No.

3255.140

Submitted to

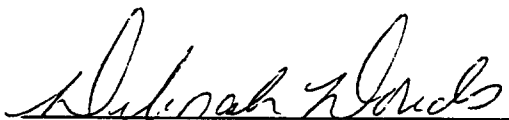
Elf Atochem North America, Inc.
2000 Market Street
Philadelphia, PA 19103

SLI Study No. 3255.140

(2)

COMPLIANCE STATEMENT

This study was conducted in compliance with the Good Laboratory Practice Standards as described by the EPA (40 CFR Part 792) and the OECD [Annex 2 C(81)30].



Deborah A. Douds, M.S.
Study Director/Author
Springborn Laboratories, Inc.

Date 12/23/97

QUALITY ASSURANCE STATEMENT

This study was inspected by the Quality Assurance Unit and reports were submitted to management and the Study Director in accordance with SLI's Standard Operating Procedures as follows:

<u>Phase</u>	<u>Date</u>
Body Weights	09/26/97
Data Audit	11/11/97
Draft Report Review	11/20/97
Final Report Review	12/23/97
Reports to Study Director and Management	11/20/97, 12/23/97

This study was conducted in compliance with the Good Laboratory Practice Standards as described by the EPA (40 CFR Part 792) and the OECD [Annex 2 C(81)30].

Richard J. Clarke
Richard J. Clarke, B.S.
Senior Quality Assurance Auditor

Date 12-23-97

Christophe W. Wilson
for Anita M. Bosau
Director of Compliance Assurance

Date 12/23/97

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SUMMARY

The potential irritant and/or corrosive effects of Sodium Hexafluoroantimonate were evaluated on the eyes of New Zealand White rabbits. One rabbit was initially dosed and received a 0.1000 g dose of the test article in the conjunctival sac of the right eye. Due to severe ocular irritation observed additional dosing was discontinued at this time. In order to more fully assess the irritant and/or corrosive potential of the test article, dosing was initiated at a later time period on an additional five rabbits. During dosing of the second set of rabbits, severe irritation was noted immediately post-dose in the first animal dosed. Due to the severe ocular response noted, the remaining four animals were not dosed. The contralateral eye of each animal remained untreated and served as a control. Test and control eyes were examined for signs of irritation for up to 21 days following dosing for the first animal and for up to 72 hours following dosing for the second animal.

Exposure to the test article produced corneal opacity in 2/2 test eyes at the 1 hour scoring interval. The corneal injury was confirmed by positive fluorescein dye retention at the 24 hour scoring interval. In both test eyes, the corneal opacity persisted until termination (following the study day 21 scoring interval for the first animal and following the 72 hour scoring interval for the remaining animal). Iritis was observed in 2/2 test eyes at the 1 hour scoring interval and resolved completely in one test eye by study day 14 and persisted in the remaining test eye to the 72 hour scoring interval. Conjunctivitis (redness, swelling and discharge) was noted in 2/2 test eyes at the 1 hour scoring interval. The conjunctival irritation persisted in one test eye to study day 21 and to the 72 hour scoring interval in the other test eye. Additional ocular findings included blanching on the nictitating membrane, corneal neovascularization, raised area on the corneal surface, blanching on the upper lid and lower lid and a red linear striation below the cornea, which were noted in 2/2, 1/2, 1/2, 2/2 and 1/2 test eyes, respectively.

Under the conditions of this test, Sodium Hexafluoroantimonate is considered to be corrosive to the ocular tissue of the rabbit.

I. INTRODUCTION

This study was performed to assess the irritant and/or corrosive effects of Sodium Hexafluoroantimonate in New Zealand White rabbits when administered by a single ocular dose. This study is intended to provide information on the potential health hazards of the test article with respect to ocular exposure. Data from this study may serve as a basis for classification and/or labeling of the test article. This study was performed at Springborn Laboratories, Inc., 553 North Broadway, Spencerville, Ohio. The protocol was signed by the Study Director on September 4, 1997 (GLP initiation date). The in-life phase of the study was initiated with test article administration on September 26, 1997 (day 0) and concluded with final scoring on October 18, 1997.

II. MATERIALS AND METHODS

A. Experimental Protocol

The protocol is included in Appendix A.

B. Test Article

The test article was received from the Sponsor and identified as follows:

Sponsor's ID	Assigned SLI ID	Physical Description	Receipt Date	Expiration Date
Sodium Hexafluoroantimonate Lot No.: PP-1-56	S97.014.3255	White crystalline powder	August 13, 1997	October 22, 1997

The test article was stored at room temperature. The Sponsor is responsible for any necessary evaluations related to the identity, strength, purity, composition, stability and method of synthesis of the test material according to 40 CFR 792.105 and OECD [Annex 2 C(81)30].

C. Retention Sample

An approximate 1 g retention sample of the test article was taken and stored at SLI at room temperature.

D. Test Article Disposition

The remaining test article was returned to the Sponsor following completion of all studies with the test article.

E. Method of Test Article Preparation

The test article was ground in a mortar with a pestle and passed through a No. 40 mesh sieve. The weight of processed test article that occupied a volume of 0.1 mL was determined and exceeded 0.1 g; therefore, 0.1000 g was utilized for dose administration.

F. Animals and Animal Husbandry

1. Description, Identification and Housing

Adult, New Zealand White rabbits were received at SLI from Myrtle's Rabbitry, Thompson Station, TN. Upon receipt, plastic ear tags displaying unique identification numbers were used to individually identify the animals. Cage cards displaying at least the study number, animal number and sex were affixed to each cage. The animals were housed individually in suspended stainless steel cages. All housing and care were based on the standards recommended by the Guide for the Care and Use of Laboratory Animals [1].

2. Environment

The animal room temperature and relative humidity ranges were 58-71°F and 30-70%, respectively. Environmental control equipment was monitored and adjusted as necessary to minimize fluctuations in the animal room environment. Light timers were set to maintain a 12-hour light/12-hour dark cycle and room ventilation was set to produce 10-15 air changes/hour. The animal room temperature and relative humidity were recorded a minimum of once daily.

3. Food

PMI Certified Rabbit Chow #5322 (Purina Mills, Inc.) was provided ad libitum to the animals throughout the study. The lot number and expiration date of each batch of diet used during the study were recorded. The feed was analyzed and certified by the supplier for nutritional components and environmental contaminants. Dietary limitations for various environmental

contaminants, including heavy metals, pesticides, polychlorinated biphenyls and total aflatoxin are set by the manufacturer. Within these limits, contaminants which may have been present were not expected to compromise the purpose of this study. Results of the dietary analyses (Certificates of Analysis) are provided by the manufacturer for each lot of diet. These are maintained by SLI.

4. Water

Municipal tap water treated by reverse osmosis was available ad libitum throughout the study. The purified water was supplied by an automatic watering system. Monitoring of the drinking water for contaminants is conducted annually by SLI and the records are available for inspection. Within generally accepted limits, contaminants which may have been present were not expected to compromise the purpose of this study. The water meets the standards specified under the EPA National Drinking Water Regulations (40 CFR, Part 141).

5. Acclimation

Upon receipt, the animals were removed randomly from the shipping cartons, examined by qualified personnel, identified with plastic ear tags and then acclimated to the laboratory conditions for a minimum of five days. The animals were observed daily for overt physical or behavioral abnormalities, general health/moribundity and mortality.

6. Animal Selection

The animals chosen for study use were arbitrarily selected from healthy stock animals to avoid potential bias. All animals received a detailed pretest observation prior to dosing. Only healthy animals were chosen for study use. Females were nulliparous and nonpregnant.

III. EXPERIMENTAL PROCEDURES

A. Preliminary Examination

On day 0 prior to dosing, both eyes of each animal provisionally selected for test use were examined macroscopically for ocular irritation with the aid of an auxiliary light source. In addition, the corneal surface was examined using fluorescein sodium dye. One drop of a fluorescein/physiological saline mixture was gently dropped onto the superior sclera of each eye. Following an approximate 15 second exposure, the

eyes were thoroughly rinsed with physiological saline. The corneal surface was then examined for dye retention under a long-wave UV light source. Animals exhibiting ocular irritation, preexisting corneal injury or fluorescein dye retention were not used on study. All animals found to be acceptable for test use were returned to their cages until dosing.

B. Dosing

A minimum of one hour after preliminary ocular examination, the test article was instilled as follows:

Group	Concentration (%)	Amount Instilled	No. of Animals	
			Males	Females
No Rinse	100	0.1000 g	1	1

The test article was instilled into the conjunctival sac of the right eye of each animal after gently pulling the lower lid away from the eye. Following instillation, the eyelids were gently held together for approximately one second in order to limit test article loss and the animal was returned to its cage. The contralateral eye remained untreated to serve as a control. One rabbit was initially dosed. Due to the severe ocular irritation observed following dosing, additional dosing was discontinued at this time. In order to more fully assess the irritant and/or corrosive potential of the test article, dosing was initiated at a later time period on an additional five rabbits. During dosing of the second set of rabbits, severe irritation was noted immediately post-dose in the first animal dosed. Therefore, the remaining four animals were not dosed.

C. Ocular Observations

The eyes were macroscopically examined with the aid of an auxiliary light source for signs of irritation at 1, 24, 48 and 72 hours and up to 21 days after dosing according to the Ocular Grading System presented in Protocol Appendix A which is based on Draize [2]. Due to the severe response noted in the second animal dosed, it was determined that additional information would not be obtained by scoring the second animal past the 72 hour scoring interval. Therefore, this animals was only scored at 1, 24, 48 and 72 hours. Following macroscopic observations at the 24 hour scoring interval, the fluorescein examination procedure was repeated on all test and control eyes and any residual test article was gently rinsed from the eye at this time (if possible) using physiological saline. If any fluorescein findings were noted at 24 hours, a fluorescein exam was conducted on the affected eyes at each subsequent interval.

D. Clinical Observations

Any unusual observations and/or mortality were recorded. General health/mortality checks were performed twice daily (in the morning and in the afternoon).

E. Body Weights

Individual body weights were obtained for each animal prior to dosing on day 0.

F. Gross Necropsy

Each animal was euthanized by an intravenous injection of sodium pentobarbital following its final observation interval. Gross necropsy examinations were not required for these animals.

G. Protocol Deviations

The animal room temperature (58-71°F) exceeded the preferred range (61-73°F) during the study. This occurrence is considered to have had no adverse effect on the outcome of this study.

IV. ANALYSIS OF DATA

The ocular irritation score for each parameter (i.e., corneal opacity x area, iritis and conjunctival redness + swelling + discharge) was multiplied by the appropriate factor (i.e., corneal injury x 5, iritis x 5, conjunctivitis x 2) and the totals added for each animal/interval. The group mean irritation score was then calculated for each scoring interval based on the number of animals initially dosed in each group.

The data will be analyzed and summarized in the report based on the definitions presented below:

1. Non Irritant - Following instillation of the test article, none of the test eyes showed a positive effect as defined in the Ocular Grading System in Protocol Appendix A.
2. Irritant - Following instillation of the test article, one or more test eyes exhibited a positive effect, but the changes were reversible.
3. Corrosive - One or more test eyes exhibited irreversible changes (i.e., necrosis or ulceration) following instillation of the test article.

V. MAINTENANCE OF RAW DATA AND RECORDS

All original paper data, the final report and magnetically encoded records were transferred to the SLI archives for a period of 10 years. The Sponsor will be contacted prior to final disposition of these items.

VI. RESULTS

A. Ocular/Clinical Observations:

Individual Data: Table 1


Individual Clinical Observations: Appendix B

Exposure to the test article produced corneal opacity in 2/2 test eyes at the 1 hour scoring interval. The corneal injury was confirmed by positive fluorescein dye retention at the 24 hour scoring interval. In both test eyes, the corneal opacity persisted until termination (following the study day 21 scoring interval for the first animal and following the 72 hour scoring interval for the remaining animal). Iritis was observed in 2/2 test eyes at the 1 hour scoring interval and resolved completely in one test eye by study day 14 and persisted in the remaining test eye to the 72 hour scoring interval. Conjunctivitis (redness, swelling and discharge) was noted in 2/2 test eyes at the 1 hour scoring interval. The conjunctival irritation persisted in one test eye to study day 21 and to the 72 hour scoring interval in the other test eye. Additional ocular findings included blanching on the nictitating membrane, corneal neovascularization, raised area on the corneal surface, blanching on the upper lid and lower lid and a red linear striation below the cornea, which were noted in 2/2, 1/2, 1/2, 2/2 and 1/2 test eyes, respectively.

No corneal opacity, iritis or conjunctivitis was observed in the control eyes.

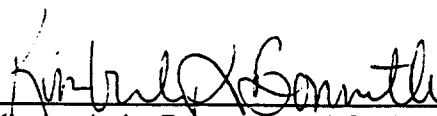
VII. CONCLUSION

Under the conditions of this test, Sodium Hexafluoroantimonate is considered to be corrosive to the ocular tissue of the rabbit.


Deborah A. Douds, M.S.
Study Director

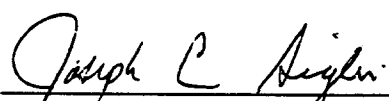
Date 12/23/97

VIII. REPORT REVIEW



Kimberly L. Bonnette, M.S., LATG
Manager of Acute Toxicology

Date 12/23/97



Joseph C. Siglin, Ph.D., DABT
Director of Toxicology

Date 12/23/97

IX. REFERENCES

1. Guide for the Care and Use of Laboratory Animals, DHHS Publication No. (NIH) 96-03, 1996.
2. Draize, J.H., Appraisal of the Safety of Chemicals in Foods, Drugs and Cosmetics, The Association of Food and Drug Officials of the United States, 49-51, 1959.

SLI STUDY NO.: 3255.140
CLIENT: ELF ATOCHEM

TABLE 1
A PRIMARY EYE IRRITATION STUDY IN RABBITS
INDIVIDUAL OCULAR IRRITATION SCORES
(NO RINSE GROUP)

PAGE 1

Animal No./Sex Body Weight (kg)	Scoring Interval	Cornea		Iris		Conjunctivae			Fluorescein		Test Eye*		Control Eye*	
		O	A	O	A	I	Ix5	R S D	(R+S+D)/2	Total	Examination	Secondary Ocular Findings	Fluorescein Examination	Secondary Ocular Findings
		3	3	4	5	1	5	2	3	2	14	64		
56105/M 2.418	1 Hour	3	3	4	5	1	5	2	3	2	14	64		
	24 Hours	3	3	4	5	1	5	2	3	3	16	66		
	48 Hours	3	3	4	5	1	5	2	3	3	16	66		
	72 Hours	3	3	4	5	1	5	2	3	2	14	64		
	7 Days	4	4	8	0	1	5	2	2	2	12	97		
	10 Days	4	4	8	0	1	5	2	2	2	12	97		
	14 Days	4	2	4	0	0	0	1	2	1	8	48		
	21 Days	4	2	4	0	0	0	2	2	1	10	50		
56175/F 2.720	1 Hour	4	4	8	0	1	5	2	1	2	10	95		
	24 Hours	4	4	8	0	1	5	2	1	2	10	95		
	48 Hours	4	4	8	0	1*	5	3	2	1	12	97		
	72 Hours	4	4	8	0	1*	5	3	2	2	14	99		

*See Protocol Appendix A for definition of codes; BCT = Blanching upper lid and lower lid; BNM = Blanching nictitating membrane; BNB = Blanching nictitating membrane.

*A red linear striation observed from approximately 11 o'clock to 7 o'clock (entire length of eye); appears to be below corneal surface but above iridal surface, confirmed by slit lamp examination.

Mean Ocular Scores

1 Hour	-	79.50
24 Hours	-	80.50
48 Hours	-	81.50
72 Hours	-	81.50
7 Days	-	97.00
10 Days	-	97.00
14 Days	-	48.00
21 Days	-	50.00

SLI Study No. 3255.140

(16)

APPENDIX A

Protocol

**A PRIMARY EYE IRRITATION STUDY IN RABBITS WITH
SODIUM HEXAFLUOROANTIMONATE**

PROTOCOL

Springborn Study No. 3255.140

Protocol No.: ELFATO/EI-1
Issue Date: February 1997

Springborn Laboratories, Inc. (SLI)
Health and Environmental Sciences
640 North Elizabeth Street
Spencerville, Ohio 45887

Deborah A. Douds, M.S.
Study Director

For

Elf Atochem North America, Inc.
2000 Market Street
Philadelphia, PA 19103

SLI Study No. 3255.140

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I. PURPOSE

The purpose of the study is to assess the irritant and/or corrosive effects of a test article in rabbits when administered by a single ocular dose. This study is intended to provide information on the potential health hazards of the test article with respect to ocular exposure. Data from this study may serve as a basis for classification and/or labeling of the test article.

II. RESPONSIBILITIES

A. Sponsor

Elf Atochem North America, Inc.
2000 Market Street
Philadelphia, PA 19103

B. Sponsor's Representative

Roy Bannister, Ph.D.
Phone: (215) 419-5875
FAX: (215) 419-5800

C. Testing Location

Springborn Laboratories, Inc.
Health and Environmental Sciences
553 North Broadway
Spencerville, OH 45887
Phone: (419) 647-4196
FAX: (419) 647-6560

D. Personnel Responsibilities

1. Deborah A. Douds, M.S.
Study Director/Assistant Manager of Acute Toxicology
2. Kimberly L. Bonnette, M.S., LATG
Alternate Contact/Manager of Acute Toxicology
3. Todd N. Merriman, B.S., LATG
Toxicologist
4. Robert C. Springborn, Ph.D.
Chairman, President and CEO
5. Malcolm Blair, Ph.D.
Vice President and Managing Director
6. Rusty E. Rush, M.S., LAT, DABT
Associate Director of Toxicology
7. J. Dale Thurman, D.V.M., M.S., DACVP
Director of Pathology
8. Anita M. Bosau
Director of Compliance Assurance

III. PROPOSED STUDY SCHEDULE

- | | |
|---------------------------------|--|
| A. Initiation of In-Life Phase: | September 1997 |
| B. Completion of In-Life Phase: | September 1997 |
| C. Audited Report Date: | 4 Weeks Following Sponsor's Approval for Study Termination |

IV. TEST ARTICLE IDENTIFICATION

A. Test Article

1. Sponsor's Identification

Sodium Hexafluoroantimonate

2. SLI Test Article Identification Number

S97.014.3255

3. Characteristics

The Sponsor is responsible for any necessary evaluations related to identity, strength, purity, composition, stability and method of synthesis of the test material according to 40 CFR 792.105 and OECD [Annex 2 C(81)30]. Any special storage conditions for the test article will be supplied by the Sponsor.

4. Handling Precautions

Safety data regarding the test article should be provided by the Sponsor [Material Safety Data Sheet (MSDS), if available]. Technical personnel are required to read this information prior to handling the test article. Any question concerning this information should be referred to the Study Director.

Additional safety and handling information may be provided by the Study Director and/or Sponsor. Minimum safety requirements include safety glasses, impervious gloves, and laboratory wear. An MSDS shall also be available for any other chemical entities utilized in the conduct of this study.

B. Retention Sample

A 1 g or 1 mL retention sample of each lot of the test article will be taken for studies with an experimental period of 4 weeks or longer. Otherwise, the Sponsor will be responsible for maintaining a retention sample of the test article.

C. Test Article Disposition

The test article will be returned to the Sponsor following completion of all studies with the test article(s) unless otherwise instructed by the Sponsor.

D. Method of Test Article Preparation

Liquids, gels and pastes are generally administered as received from the Sponsor. Solids and powders are generally ground and sieved prior to test use. This may be accomplished by grinding the material in a mortar and pestle and passing the material through a No. 40 mesh sieve. The weight of processed test article that occupies a volume of 0.1 mL will be determined by measuring a convenient volume (at least 2 mL) of the powder in a suitable volumetric container. The powder will be gently compacted by tapping the measuring container. The test article dose per eye will then be calculated (weight equivalent of 0.1 mL, not to exceed 0.1 g). The test article will be prepared and/or dispensed fresh on the day of dosing. The method of preparation will be documented in the raw data and presented in the final report.

V. TEST SYSTEM

A. Justification of the Test System

1. The rabbit is the preferred species for primary eye irritation testing by various U.S. and International regulatory agencies.
2. The New Zealand White rabbit has been shown to be sensitive to the irritant/corrosive effects of a variety of drugs and chemicals. Therefore, this species and strain is a reasonable alternative to larger mammals for primary eye irritation testing of drugs and chemicals for human safety assessment.
3. The New Zealand White rabbit has been used extensively for eye irritation testing. Thus, data from this study may be compared and contrasted to other studies performed in New Zealand White rabbits.
4. Historical information concerning New Zealand White rabbits is available at SLI and in the published literature.

5. Healthy, outbred New Zealand White rabbits may be obtained from reliable, USDA approved and regulated suppliers.
6. The laboratory rabbit may be safely handled and manipulated by trained technical personnel.

SLI has conducted literature searches through Medline, Toxline and Bioethics and there are no generally accepted validated alternatives to this test. In a 1989 position paper prepared by the Animals in Research Committee of The Society of Toxicology (SOT) and approved by the SOT Council, it was concluded that "none of these proposed models are yet validated or evaluated for a broad range of chemical moieties, and none can be relied upon to provide the scientific reliability or predictive accuracy which would be required for a new test for regulatory of legal acceptability" [1].

B. Justification of the Route of Exposure and Number of Animals

1. Ocular administration of the test substance was selected since this is a potential route of human exposure.
2. Since New Zealand White rabbits have no pigment and have an easily accessible ocular area, substances may be accurately instilled and any resulting effects easily observed.
3. The number of animals used on this study will be consistent with the guidelines published by a number of U.S. and International regulatory agencies including EPA-FIFRA, EPA-TSCA, EEC, OECD and MAFF.

C. Description

1. Species

Rabbit

2. Strain

New Zealand White

3. Source

Myrtle's Rabbitry or another USDA approved supplier

4. Age and Body Weight Range

Adult, approximately 2.0 to 3.5 kg (prior to dosing on day 0).

5. Number of Animals/Sex on Study

6 rabbit test (males and/or females)

D. Method of Identification

Plastic ear tags displaying unique identification numbers will be used to individually identify the animals. The cage cards will display at least the study number, animal number, and sex and will be affixed to each cage.

E. Animal Husbandry

1. Housing

The animals will be housed individually in suspended stainless steel cages. All housing and care will be based on the standards recommended by the Guide for the Care and Use of Laboratory Animals [2].

2. Environment

The environmental conditions for the animal room will be set to maintain room temperature and relative humidity ranges of $67 \pm 6^{\circ}\text{F}$ and $50 \pm 20\%$, respectively. Environmental control equipment will be monitored and adjusted as necessary to minimize fluctuations in the animal room environment. Light timers will be set to maintain a 12-hour light/12-hour dark cycle and the room ventilation will be set to produce 10-15 air changes/hour. The room temperature and relative humidity will be recorded a minimum of once daily.

3. Food

PMI Certified Rabbit Chow #5322 (Purina Mills, Inc.) will be provided ad libitum to the animals throughout the study. The lot number and expiration date of each batch of diet used during the study will be recorded. The feed is analyzed and certified by the supplier for nutritional components and environmental contaminants. Dietary limitations for various environmental contaminants, including heavy metals, pesticides, polychlorinated biphenyls and total aflatoxin are set by the manufacturer. Within these limits, there are no contaminants reasonably expected in the diet which would interfere with the conduct of the study. Results of the dietary analyses (Certificates of Analysis) are provided by the manufacturer for each lot of diet. These will be maintained in the laboratory records. Feed that is outside the ranges set for the above mentioned criteria will not be utilized by the testing facility.

4. Water

Municipal tap water following treatment by reverse osmosis will be available ad libitum throughout the study. The purified water will be supplied by an automatic watering system. Monitoring of the drinking water for contaminants will be conducted annually by the testing laboratory and the records will be available for inspection. Levels of contaminants which may be present are not expected to compromise the purpose of the study. The water meets the standards specified under the EPA National Drinking Water Regulations (40 CFR, Part 141).

F. Acclimation

Upon receipt, the animals will be removed randomly from the shipping cartons, examined by qualified personnel, identified with plastic ear tags, and then acclimated to the laboratory conditions for a minimum of 5 days. The animals will be observed daily for overt physical or behavioral abnormalities, general health/moribundity and mortality.

G. Animal Selection

The animals chosen for study use will be arbitrarily selected from healthy stock animals to avoid potential bias. All animals will receive a detailed pretest observation prior to dosing. Only healthy animals will be chosen for study use. Females will be nulliparous and nonpregnant.

VI. EXPERIMENTAL DESIGN AND PROCEDURES

A. Study Group Design

A six rabbit test will be performed. Materials which are determined by the Sponsor to be strong acids ($\text{pH} \leq 2.0$), strong alkalis ($\text{pH} \geq 11.5$) or a material which produces severe dermal irritation need not be tested in a full number of animals due to their predictive corrosive properties. However, at the request of the Sponsor, these materials will be administered to one animal. If no corrosive response is seen during the first 72 hours, the material will be tested on the remaining five animals.

B. Preliminary Examination

On day 0 prior to dosing, both eyes of each animal provisionally selected for test use will be examined macroscopically for ocular irritation with the aid of an auxiliary light source. In addition, the corneal surface will be examined using fluorescein sodium dye. One drop of a fluorescein/physiological saline mixture will be gently dropped onto the superior sclera of each eye. Following an approximate 15 second exposure, the eyes will be thoroughly rinsed with physiological saline. The corneal surface will then be examined for dye retention under a long-wave UV light source. Animals exhibiting ocular irritation, preexisting corneal injury or fluorescein dye retention will not be used on study. All animals found to be acceptable for test use will be returned to their cages until dosing.

C. Dosing

A minimum of one hour after preliminary ocular examination, the test article will be instilled into the conjunctival sac of the right eye of each animal after gently pulling the lower lid away from the eye. Liquids, gels and pastes will be administered at a volume of 0.1 mL. Solids and powders will be administered at a weight equivalent to 0.1 mL volume, not to exceed 0.1 g. Following instillation, the eyelids will be gently held together for approximately one second in order to limit test article loss and the animal returned to its cage. The contralateral eye will remain untreated to serve as a control. Following dosing, the Study Director will be notified by the technician if severe local reactions occur or if the animals exhibit overt clinical indications of pain/distress immediately post-dose. If such is noted, the Sponsor will be contacted to see if the animals should be humanely euthanized.

D. Body Weights

Individual body weights will be obtained for each animal prior to dosing on day 0.

E. Ocular Observations

The eyes will be macroscopically examined with the aid of an auxiliary light source for signs of irritation at 1, 24, 48 and 72 hours after dosing according to the Ocular Grading System presented in Protocol Appendix A which is based on Draize [3]. At the discretion of the Study Director, a biomicroscopic slit-lamp may be utilized to further examine and clarify ocular lesions. Following macroscopic observations at the 24 hour scoring interval, the fluorescein examination procedure will be repeated on all test and control eyes and any residual test article should be gently rinsed from the eye at this time (if possible) using physiological saline. If any fluorescein findings are noted at 24 hours, a fluorescein exam will be conducted on the affected eyes at each subsequent interval until a negative response is obtained and/or until all corneal opacity has cleared, or as directed by the Study Director. If there is no evidence of treatment-related ocular irritation at the 72 hour scoring interval, the study will be terminated. If ocular irritation persists in any test eye, the observation period may be extended for the affected animals (scored on days 7, 10, 14 and 21). Animals requiring an extended observation period will remain on test until the irritation has resolved, permanent injury is evident or the Study Director/Sponsor determines that additional scoring intervals are unnecessary.

F. Clinical Observations

Any unusual observations and mortality will be recorded. General health/mortality checks will be performed twice daily (in the morning and in the afternoon).

G. Unscheduled Deaths and Euthanasia

Any animals dying or euthanized for cause during the study period will be necropsied. The animals will be euthanized by an intravenous injection of sodium pentobarbital. Body cavities (cranial, thoracic, abdominal and pelvic) will be opened and examined. No tissues will be retained.

H. Scheduled Euthanasia

Each surviving animal will be euthanized by an intravenous injection of sodium pentobarbital following its final observation interval. A gross necropsy examination will not be required for surviving animals.

VII. PROTOCOL AMENDMENT

Alterations to this protocol may be made as the study progresses. No changes in the protocol will be made without the specific consent of the Sponsor's Representative. A protocol amendment will be prepared and signed by the Study Director, SLI Quality Assurance and Sponsor's Representative for any such changes.

VIII. DATA REPORTING

One unbound copy of the draft report and two copies of the final report (one bound and one unbound) will be submitted to the Sponsor. The final report will include all information necessary to provide a complete and accurate description and evaluation of the experimental procedures and results.

The report will include at least the following information and data:

- Table of Contents
- Regulatory Compliance
- Summary
- Introduction
- Experimental Design and Test Procedures
- Presentation and Discussion of Results
- Conclusion
- References
- Data Tables
- Protocol and Amendments
- SLI Personnel Responsibilities

IX. ANALYSIS OF DATA

For each group, the ocular irritation score for each parameter (i.e., corneal opacity x area, iritis and conjunctival redness + swelling + discharge) will be multiplied by the appropriate factor (i.e., corneal injury x 5, iritis x 5, conjunctivitis x 2) and the totals added for each animal/interval. The group mean irritation score will then be calculated for each scoring interval based on the number of animals initially dosed in each group. If an animal dies during the study, the total animals in that group will be reduced (by the number of animals dead) for each subsequent scoring interval for the purpose of calculating the mean ocular irritation score for each interval.

The data will be analyzed and summarized in the report based on the definitions presented below:

1. Non Irritant - Following instillation of the test article, none of the test eyes show a positive effect as defined in the Ocular Grading System in Protocol Appendix A.
2. Irritant - Following instillation of the test article, one or more test eyes exhibit a positive effect, but the changes are reversible.
3. Corrosive - One or more test eyes exhibit irreversible changes (i.e., necrosis or ulceration) following instillation of the test article.

X. MAINTENANCE OF RAW DATA, RECORDS AND SPECIMENS

All original data, magnetically encoded records, specimens and reports from this study are the property of the Sponsor. These materials shall be available at SLI to facilitate auditing of the study during its progress and prior to acceptance of the final report. All original paper data, the final report, magnetically encoded records, and any specimens will be transferred to the SLI archives for a period of 10 years. The Sponsor will be contacted prior to the final disposition of these items.

XI. REGULATORY COMPLIANCE

This study may be submitted to and will be conducted in accordance with the EPA-TSCA [4], OECD [5] and EEC [6] guidelines and the principles of the Good Laboratory Practice Standards by the EPA (40 CFR Part 792) and the OECD [Annex 2 C(81)30].

XII. QUALITY ASSURANCE

The study will be inspected at least once during the in-life phase by the Springborn Laboratories, Inc., Quality Assurance Unit to assure compliance with Good Laboratory Practice Standards, SLI's Standard Operating Procedures and for conformance with the protocol and protocol amendments. The final report will be audited prior to submission to the Sponsor to ensure that it completely and accurately describes the test procedures and results of the study.

XIII. USDA ANIMAL WELFARE COMPLIANCE STATEMENT

This study will comply with all applicable sections of the Final Rules of the Animal Welfare Act regulations (9 CFR) and the Public Health Service Policy on Humane Care and Use of Laboratory Animals (OPRR, NIH, 1986). Wherever possible, procedures used in this study have been designed to avoid or minimize discomfort, distress and pain to animals. All methods are described in this study protocol or in written laboratory standard operating procedures. These procedures are based on the most currently available technologies concerning proper laboratory animal use and management. Methods of euthanasia used during this study are in conformance with the above referenced regulations and the American Veterinary Medical Association Panel on Euthanasia [7]. This protocol has been reviewed and approved by Springborn Laboratories, Inc. Institutional Animal Care and Use Committee (IACUC) for a maximum of 18 animals. Prior IACUC approval will be obtained for repeated studies.

This study is being conducted to evaluate potential irritant effects of the test article and potential reversibility of such effects. Following dosing, the Study Director will be notified by the technician if severe local reactions occur or if the animals exhibit overt clinical indications of pain/distress post-dose or if delayed severe ocular or clinical changes subsequently develop. If severe reactions are noted, the Study Director will contact the Facility Veterinarian and Sponsor to consider an appropriate course of action. In the event that the Sponsor cannot be contacted, the Study Director and Facility Veterinarian may authorize treatment or euthanasia of the animals. In general, the ocular tissue will not be anesthetized prior to or following dosing since these substances have been shown to inhibit the blink and/or tear response which may alter the irritation response. Furthermore, anesthetic agents may interact with and/or dilute the test article and thereby alter the experimental results. However, if it is suspected that the test article may induce more than transient pain/distress based on existing information, preanesthesia will be considered. In such circumstances, the Study Director and/or Facility Veterinarian will consult with the Sponsor to devise an appropriate study plan.

XIV. DECLARATION OF INTENT

This study will be listed on the SLI Quality Assurance Master Schedule for the EPA.

XV. GENERIC PROTOCOL APPROVAL

The Sponsor's signature below documents that there are no acceptable non-animal alternatives for this study, and that since this study is required by the relevant supervising government agency, it does not unnecessarily duplicate any previous experiments.

Kimberly L. Bonnette
Kimberly L. Bonnette, M.S., LATG
Manager of Acute Toxicology (SLI)

Date: 3/4/97

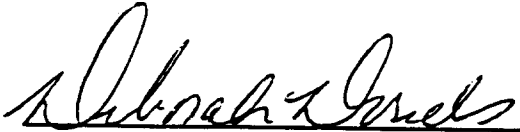
Christopher W. Wilson
Christopher W. Wilson, B.S.
Supervisor of Quality Assurance (SLI)

Date: 3/5/97

Roy B. Bristow
Roy Bristow, Ph.D.
Sponsor's Representative
(Principal Investigator)

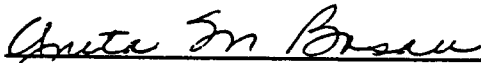
Date: 2/23/97

XVI. STUDY SPECIFIC PROTOCOL APPROVAL



Deborah A. Douds, M.S.
Study Director (SLI)

Date: 9/4/97



Quality Assurance Unit (SLI)

Date: 9/4/97

XVI. REFERENCES

1. SOT Position Paper, "Comments on the LD50 and Acute Eye and Skin Irritation Tests, Fundamental and Applied Toxicology 13, 621-623, 1993.
2. Guide for the Care and Use of Laboratory Animals, DHHS Publication No. (NIH) 96-03, 1996.
3. Draize, J.H., Appraisal of the Safety of Chemicals in Foods, Drugs and Cosmetics, The Association of Food and Drug Officials of the United States, 49-51, 1959.
4. Toxic Substances Control Act Test Guidelines, 40 CFR Part 798, Subpart E, Section 798.4500, July 1, 1992.
5. OECD Guidelines for the Testing of Chemicals, Section 4, Health Effects, Subsection 405, February 24, 1987.
6. The EEC Guidelines Part B: Method for the Determination of Toxicity, No. L 383 A/127, B.5, December 29, 1992.
7. 1993 Report of the American Veterinary Medical Assoc. Panel on Euthanasia, JAVMA, Vol. 202, No. 2, pp. 229-249, January 15, 1993.

PROTOCOL APPENDIX A

Ocular Grading System

(O) CORNEAL OPACITY--DEGREE OF DENSITY (AREA MOST DENSE TAKEN FOR READING)	
OBSERVATION	CODE
No ulceration or opacity	0
Scattered or diffuse areas of opacity (other than slight dulling of normal luster), details of iris clearly visible	1*
Easily discernible translucent area, details of iris slightly obscured	2*
Nacreous (opalescent) area, no details of iris visible, size of pupil barely discernible	3*
Opaque cornea, iris not discernible through opacity	4*

(A) AREA OF CORNEA INVOLVED (TOTAL AREA EXHIBITING ANY OPACITY, REGARDLESS OF DEGREE)	
OBSERVATION	CODE
No ulceration or opacity	0
One quarter (or less) but not zero	1
Greater than one quarter, but less than half	2
Greater than half, but less than three quarters	3
Greater than three quarters, up to whole area	4

Cornea Score = O x A x 5

Total Maximum = 80

(I) IRITIS	
OBSERVATION	CODE
Normal	0
Markedly deepened rugae (folds above normal), congestion, swelling, moderate circumcorneal hyperemia or injection, any or all of these or combination of any thereof, iris is still reacting to light (sluggish reaction is positive)	1*
No reaction to light, hemorrhage, gross destruction (any or all of these)	2*

Iris Score = I x 5

Total Maximum = 10

*Positive Effect.

PROTOCOL APPENDIX A--(Continued)

Ocular Grading System

(R) CONJUNCTIVAL REDNESS (REFERS TO PALPEBRAL AND BULBAR CONJUNCTIVAE EXCLUDING CORNEA AND IRIS)	
OBSERVATION	CODE
Blood vessels normal	0
Some blood vessels definitely hyperemic (injected) above normal (slight erythema)	1
Diffuse, crimson color, individual vessels not easily discernible (moderate erythema)	2*
Diffuse beefy red (marked erythema)	3*

(S) CONJUNCTIVAL SWELLING (LIDS AND/OR NICTATING MEMBRANE)	
OBSERVATION	CODE
No swelling	0
Any swelling above normal (includes nictitating membrane, slightly swollen)	1
Obvious swelling with partial eversion of lids	2*
Swelling with lids about half closed	3*
Swelling with lids more than half closed	4*

(D) CONJUNCTIVAL DISCHARGE	
OBSERVATION	CODE
No discharge	0
Any amount different from normal (does not include small amounts observed in inner canthus of normal animals)	1
Discharge with moistening of the lids and hairs just adjacent to lids	2
Discharge with moistening of the lids and hairs and considerable area around the eye	3

Conjunctival Score = (R + S + D) x 2

Total Maximum = 20

*Positive Effect.

PROTOCOL APPENDIX A—(Continued)

Ocular Grading System

CORNEAL NEOVASCULARIZATION		
OBSERVATION	CODE	DEFINITION
Neovascularization - Very Slight	VAS-1	Total area of vascularized corneal tissue is < 10% of corneal surface
Neovascularization - Mild	VAS-2	Total area of vascularized corneal tissue is > 10% but < 25% of corneal surface
Neovascularization - Moderate	VAS-3	Total area of vascularized corneal tissue is > 25% but < 50% of corneal surface
Neovascularization - Severe	VAS-4	Total area of vascularized corneal tissue is > 50% of corneal surface

SECONDARY OCULAR FINDINGS		
OBSERVATION	CODE	DEFINITION
Sloughing of the corneal epithelium	SCE	Corneal epithelial tissue is observed to be peeling off the corneal surface.
Corneal bulging	CB	The entire corneal surface appears to be protruding outward further than normal.
Slight dulling of normal luster of the cornea	SDL	The normal shiny surface of the cornea has a slightly dulled appearance.
Raised area on the corneal surface	RAC	A defined area on the corneal surface that is raised above the rest of the cornea. This area is generally associated with neovascularization and has a off-white to yellow color.
Corneal edema	CE	The cornea has a swollen appearance.
Test article present in eye	TAE	Apparent residual test article is observed on the eye or in the conjunctival sac/inner canthus.
Observation confirmed by slit lamp	OCS	A slit lamp examination was performed to confirm the initial observation.
Corneal mineralization	CM	Small white or off-white crystals that are observed in the corneal tissue.

PROTOCOL APPENDIX A--(Continued)

Ocular Grading System

FLUORESCEIN EXAMINATION OF CORNEA	
OBSERVATION	CODE
<u>Fluorescein Dye Retention</u> Fluorescein dye retention associated with the area of corneal opacity Fluorescein dye retention is not associated with any other finding	FAO FNF
<u>Negative Results</u> No fluorescein retention is observed	(-)
<u>Secondary Ocular Findings</u> Superficial mechanical abrasion to the cornea observed during the fluorescein examination period Fine stippling on the cornea observed during the fluorescein examination procedure	MI ST

POST-DOSE CLINICAL OBSERVATIONS	
OBSERVATION	CODE
Animal vocalized following dosing	VOC
Animal excessively pawed test eye following dosing	PAW
Animal exhibited excessive hyperactivity following dosing	HYP
Animal exhibited excessive head tilt following dosing	HT
Animal exhibited excessive squinting of test eye following dosing	SQ

Any additional findings will be noted in the raw data and in the final report.

APPENDIX B

Individual Clinical Observations

SLI STUDY NO.: 3255.140
CLIENT: ELF ATOCHEM

A PRIMARY EYE IRRITATION STUDY IN RABBITS
INDIVIDUAL CLINICAL OBSERVATIONS
(POSITIVE FINDINGS)

PAGE 1

Animal No./Sex	Clinical Observations
56105/M	Animal vocalized following dosing: Day 0 Hairloss - slight, around right eye: Day 14
56175/F	Animal vocalized following dosing: Day 0

APPENDIX C

SLI Personnel Responsibilities

SLI PERSONNEL RESPONSIBILITIES

Deborah A. Douds, M.S.

Study Director/Assistant Manager of
Acute Toxicology

Kimberly L. Bonnette, M.S., LATG

Alternate Contact/Manager of Acute
Toxicology

Malcolm Blair, Ph.D.

Vice President and Managing Director

Joseph C. Siglin, Ph.D., DABT

Director of Toxicology

Pamela S. Smith, ALAT

Unit Leader

Delores P. Knippen

Supervisor of Pharmacy

Jan K. Severt, B.S., ALAT

Supervisor of Acute Report Preparation

Anita M. Bosau

Director of Compliance Assurance

Christopher W. Wilson, B.S.

Supervisor of Acute Quality Assurance

Deanna M. Talerico

Supervisor of Nonacute Quality
Assurance

J. Dale Thurman, D.V.M., M.S.,
DACVP

Director of Pathology

**A PRIMARY EYE IRRITATION STUDY IN RABBITS
WITH LITHIUM TETRAFLUOROBORATE**

AMENDED FINAL REPORT

Author

Deborah A. Douds, M.S.

Original Study Completion Date

December 29, 1997

Amended Study Completion Date

January 12, 1998

Performing Laboratory

Springborn Laboratories, Inc. (SLI)
Health and Environmental Sciences
640 North Elizabeth Street
Spencerville, Ohio 45887

SLI Study No.

3255.142

Submitted to

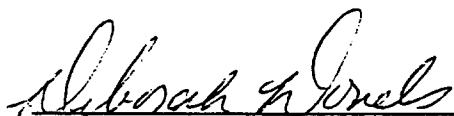
Elf Atochem North America, Inc.
2000 Market Street
Philadelphia, PA 19103

SLI Study No. 3255.142

(2)

COMPLIANCE STATEMENT

This study was conducted in compliance with the Good Laboratory Practice Standards as described by the EPA (40 CFR Part 792) and the OECD [Annex 2 C(81)30].



Deborah A. Douds, M.S.
Study Director/Author
Springborn Laboratories, Inc.

Date 12/29/97

QUALITY ASSURANCE STATEMENT

This study was inspected by the Quality Assurance Unit and reports were submitted to management and the Study Director in accordance with SLI's Standard Operating Procedures as follows:

<u>Phase</u>	<u>Date</u>
Ocular Observations	10/03/97
Data Audit	11/10/97
Draft Report Review	11/20/97
Final Report Review	12/29/97
Amended Final Report Review	01/12/98
Reports to Study Director and Management	11/20/97, 12/29/97, 01/12/98

This study was conducted in compliance with the Good Laboratory Practice Standards as described by the EPA (40 CFR Part 792) and the OECD [Annex 2 C(81)30].

Richard J. Clarke
Richard J. Clarke, B.S.
Senior Quality Assurance Auditor

Date 1-12-98

Anita M. Bosau
Anita M. Bosau
Director of Compliance Assurance

Date 1/12/98

The following page has been incorporated into this report.

Page No.	Revision	Reason for Change
5	Added page numbers to List of Tables and Appendices.	Inadvertently omitted at time of report finalization.

Deborah A. Douds
Deborah A. Douds, M.S.
Study Director

Date 1/12/98

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SUMMARY

The potential irritant and/or corrosive effects of Lithium Tetrafluoroborate were evaluated on the eyes of New Zealand White rabbits. Each of six rabbits received a 0.1000 g dose (0.1 mL weight equivalent) of the test article in the conjunctival sac of the right eye. The contralateral eye of each animal remained untreated and served as a control. Test and control eyes were examined for signs of irritation for up to 21 days following dosing.

Exposure to the test article produced corneal opacity in 6/6 test eyes at the 1 hour scoring interval. The corneal injury was confirmed by positive fluorescein dye retention at the 24 hour scoring interval. The corneal opacity resolved in 1/6 test eyes by study day 7. The corneal opacity remained in 5/6 test eyes at study termination (day 21). Iritis was observed in 6/6 test eyes at the 1 hour scoring interval which resolved completely in all test eyes by study day 21. Conjunctivitis (redness, swelling and discharge) was noted in 6/6 test eyes at the 1 hour scoring interval. The conjunctival irritation did not resolve completely in any test eyes by study day termination. Additional ocular findings noted during the study included blanching on the nictating membrane (6/6 test eyes), blanching on the lower lid of the conjunctival tissue (6/6 test eyes), apparent pseudopterygium (a portion of the conjunctival tissue growing over the cornea, most likely a conjunctival scar - 1/6 test eyes), corneal neovascularization (6/6 test eyes), raised area(s) on the cornea (2/6 test eyes) and sloughing of the corneal epithelium (4/6 test eyes). Vocalization was observed in five animals immediately following dosing.

Under the conditions of this test, Lithium Tetrafluoroborate is considered to be corrosive to the ocular tissue of the rabbit.

I. INTRODUCTION

This study was performed to assess the irritant and/or corrosive effects of Lithium Tetrafluoroborate in New Zealand White rabbits when administered by a single ocular dose. This study is intended to provide information on the potential health hazards of the test article with respect to ocular exposure. Data from this study may serve as a basis for classification and/or labeling of the test article. This study was performed at Springborn Laboratories, Inc., 553 North Broadway, Spencerville, Ohio. The protocol was signed by the Study Director on September 4, 1997 (GLP initiation date). The in-life phase of the study was initiated with test article administration on October 3, 1997 (day 0) and concluded with final scoring on October 29, 1997.

II. MATERIALS AND METHODS

A. Experimental Protocol

The protocol is included in Appendix A.

B. Test Article

The test article was received from the Sponsor and identified as follows:

Sponsor's ID	Assigned SLI ID	Physical Description	Receipt Date	Expiration Date
Lithium Tetrafluoroborate Lot No.: AQ-2-166(F)	S97.015.3255	White powder	August 13, 1997	February 3, 1998

The test article was stored at room temperature. The Sponsor is responsible for any necessary evaluations related to the identity, strength, purity, composition, stability and method of synthesis of the test material according to 40 CFR 792.105 and OECD [Annex 2 C(81)30]. A Certificate of Analysis for the test article was provided by the Sponsor and is presented in Appendix B.

C. Retention Sample

The Sponsor was responsible for maintaining a retention sample of the test article.

D. Test Article Disposition

The remaining test article was returned to the Sponsor following completion of all studies with the test article.

E. Method of Test Article Preparation

The test article was ground in a mortar with a pestle and passed through a No. 40 mesh sieve. The weight of processed test article that occupied a volume of 0.1 mL was then determined. Since this weight exceeded 0.1 g, 0.1000 g was utilized for dose administration.

F. Animals and Animal Husbandry

1. Description, Identification and Housing

Adult, New Zealand White rabbits were received at SLI from Myrtle's Rabbitry, Thompson Station, TN. Upon receipt, plastic ear tags displaying unique identification numbers were used to individually identify the animals. Cage cards displaying at least the study number, animal number and sex were affixed to each cage. The animals were housed individually in suspended stainless steel cages. All housing and care were based on the standards recommended by the Guide for the Care and Use of Laboratory Animals [1].

2. Environment

The animal room temperature and relative humidity ranges were 58-71°F and 26-79%, respectively. Environmental control equipment was monitored and adjusted as necessary to minimize fluctuations in the animal room environment. Light timers were set to maintain a 12-hour light/12-hour dark cycle and room ventilation was set to produce 10-15 air changes/hour. The animal room temperature and relative humidity were recorded a minimum of once daily.

3. Food

PMI Certified Rabbit Chow #5322 (Purina Mills, Inc.) was provided ad libitum to the animals throughout the study. The lot number and expiration date of each batch of diet used during the study were recorded. The feed was analyzed and certified by the supplier for nutritional components and environmental contaminants. Dietary limitations for various environmental contaminants, including heavy metals, pesticides, polychlorinated biphenyls and total aflatoxin are set by the manufacturer. Within these limits, contaminants which may have been present were not expected.

to compromise the purpose of this study. Results of the dietary analyses (Certificates of Analysis) are provided by the manufacturer for each lot of diet. These are maintained by SLI.

4. Water

Municipal tap water treated by reverse osmosis was available ad libitum throughout the study. The purified water was supplied by an automatic watering system. Monitoring of the drinking water for contaminants is conducted annually by SLI and the records are available for inspection. Within generally accepted limits, contaminants which may have been present were not expected to compromise the purpose of this study. The water meets the standards specified under the EPA National Drinking Water Regulations (40 CFR, Part 141).

5. Acclimation

Upon receipt, the animals were removed randomly from the shipping cartons, examined by qualified personnel, identified with plastic ear tags and then acclimated to the laboratory conditions for a minimum of five days. The animals were observed daily for overt physical or behavioral abnormalities, general health/moribundity and mortality.

6. Animal Selection

The animals chosen for study use were arbitrarily selected from healthy stock animals to avoid potential bias. All animals received a detailed pretest observation prior to dosing. Only healthy animals were chosen for study use. Females were nulliparous and nonpregnant.

III. EXPERIMENTAL PROCEDURES

A. Preliminary Examination

On day 0 prior to dosing, both eyes of each animal provisionally selected for test use were examined macroscopically for ocular irritation with the aid of an auxiliary light source. In addition, the corneal surface was examined using fluorescein sodium dye. One drop of a fluorescein/physiological saline mixture was gently dropped onto the superior sclera of each eye. Following an approximate 15 second exposure, the eyes were thoroughly rinsed with physiological saline. The corneal surface was then examined for dye retention under a long-wave UV light source. Animals exhibiting ocular irritation, preexisting corneal injury or fluorescein dye retention were not used on

study. All animals found to be acceptable for test use were returned to their cages until dosing.

B. Dosing

A minimum of one hour after preliminary ocular examination, the test article was instilled as follows:

Group	Concentration (%)	Amount Instilled	No. of Animals	
			Males	Females
No Rinse	100	0.1000 g	2	4

The test article was instilled into the conjunctival sac of the right eye of each animal after gently pulling the lower lid away from the eye. Following instillation, the eyelids were gently held together for approximately one second in order to limit test article loss and the animal was returned to its cage. The contralateral eye remained untreated to serve as a control.

Initially, one animal was dosed. Since no adverse ocular response was noted following the 72 hour scoring interval for this animal, the remaining five animals were dosed as previously described.

C. Ocular Observations

The eyes were macroscopically examined with the aid of an auxiliary light source for signs of irritation at 1, 24, 48 and 72 hours and up to 21 days after dosing according to the Ocular Grading System presented in Protocol Appendix A which is based on Draize [2]. Following macroscopic observations at the 24 hour scoring interval, the fluorescein examination procedure was repeated on all test and control eyes and any residual test article was gently rinsed from the eye at this time (if possible) using physiological saline. If any fluorescein findings were noted at 24 hours, a fluorescein exam was conducted on the affected eyes at each subsequent interval until a negative response was obtained.

D. Clinical Observations

Any unusual observations and/or mortality were recorded. General health/mortality checks were performed twice daily (in the morning and in the afternoon).

E. Body Weights

Individual body weights were obtained for each animal prior to dosing on day 0.

F. Scheduled Euthanasia

Each animal was euthanized by an intravenous injection of sodium pentobarbital following its final observation interval. Gross necropsy examinations were not required for these animals.

G. Protocol Deviations

The temperature and relative humidity of the animal room (58-71°F and 26-79%, respectively) exceeded the preferred ranges (61-73°F and 30-70%, respectively) during this study. These occurrences are considered to have had no adverse effect on the outcome of this study.

IV. ANALYSIS OF DATA

The ocular irritation score for each parameter (i.e., corneal opacity x area, iritis and conjunctival redness + swelling + discharge) was multiplied by the appropriate factor (i.e., corneal injury x 5, iritis x 5, conjunctivitis x 2) and the totals added for each animal/interval. The group mean irritation score was then calculated for each scoring interval based on the number of animals initially dosed in each group.

The data will be analyzed and summarized in the report based on the definitions presented below:

1. Nonirritant - Following instillation of the test article, none of the test eyes showed a positive effect as defined in the Ocular Grading System in Protocol Appendix A.
2. Irritant - Following instillation of the test article, one or more test eyes exhibited a positive effect, but the changes were reversible.
3. Corrosive - One or more test eyes exhibited irreversible changes (i.e., necrosis or ulceration) following instillation of the test article.

V. MAINTENANCE OF RAW DATA AND RECORDS

All original paper data, the final report and magnetically encoded records were transferred to the SLI archives for a period of 10 years. The Sponsor will be contacted prior to final disposition of these items.

VI. RESULTS

A. Ocular Observations:

Individual Data: Table 1

Individual Clinical Observations: Appendix C

Exposure to the test article produced corneal opacity in 6/6 test eyes at the 1 hour scoring interval. The corneal injury was confirmed by positive fluorescein dye retention at the 24 hour scoring interval. The corneal opacity resolved in 1/6 test eyes by study day 7. The corneal opacity remained in 5/6 test eyes at study termination (day 21). Iritis was observed in 6/6 test eyes at the 1 hour scoring interval which resolved completely in all test eyes by study day 21. Conjunctivitis (redness, swelling and discharge) was noted in 6/6 test eyes at the 1 hour scoring interval. The conjunctival irritation did not resolve completely in any test eyes by study day termination. Additional ocular findings noted during the study included blanching on the nictating membrane (6/6 test eyes), blanching on the lower lid of the conjunctival tissue (6/6 test eyes), apparent pseudopterygium (a portion of the conjunctival tissue growing over the cornea, most likely a conjunctival scar - 1/6 test eyes), corneal neovascularization (6/6 test eyes), raised area(s) on the cornea (2/6 test eyes) and sloughing of the corneal epithelium (4/6 test eyes). Vocalization was observed in five animals immediately following dosing.

No corneal opacity, iritis or conjunctivitis was observed in the control eyes.

VII. CONCLUSION

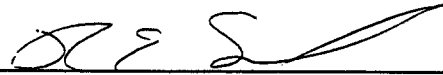
Under the conditions of this test, Lithium Tetrafluoroborate is considered to be corrosive to the ocular tissue of the rabbit.



Deborah A. Douds, M.S.
Study Director

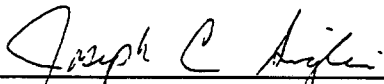
Date 12/29/97

VIII. REPORT REVIEW



for Kimberly L. Bonnette, M.S., LATG
Manager of Acute Toxicology

Date 12-29-97



Joseph C. Siglin, Ph.D., DABT
Director of Toxicology

Date 12/29/97

IX. REFERENCES

1. Guide for the Care and Use of Laboratory Animals, DHHS Publication No. (NIH) 96-03, 1996.
2. Draize, J.H., Appraisal of the Safety of Chemicals in Foods, Drugs and Cosmetics, The Association of Food and Drug Officials of the United States, 49-51, 1959.

TABLE 1
A PRIMARY EYE IRRITATION STUDY IN RABBITS
INDIVIDUAL OCULAR IRRITATION SCORES

Animal No./Sex Body Weight (kg)	Scoring Interval	Cornea		Iris		Conjunctivae			Fluorescein		Test Eye*		Control Eye*			
		O	A	Ox	Ax5	I	Ix5	R	S	D	(R+S+D)/2	Total	Exam	Secondary Ocular Findings	Fluorescein Exam	Secondary Ocular Findings
55983/M 3.433	1 Hour	3	2	30	1	5	2	2	1	10	45		BLN,BLT			
	24 Hours	3	2	30	1	5	3	2	2	14	49		BLN,BLT			
	48 Hours	3	2	30	1	5	3	2	2	14	49		BLN,BLT			[-]
	72 Hours	3	2	30	1	5	3	2	2	14	49		BLN,BLT,VAS-1			
	7 Days	4	3	60	1	5	3	2	2	14	79		BLN,BLT,VAS-3			
	10 Days	3	2	30	0	0	2	2	0	8	38		BLN,BLT,VAS-2			
	14 Days	2	1	10	0	0	1	2	0	6	16		BLN,VAS-1			
	21 Days	1	1	5	0	0	0	1	0	2	7		VAS-1,RAC			
56192/F 2.509	1 Hour	3	3	45	1	5	2	2	2	12	62		BLN,BLT			
	24 Hours	2	4	40	1	5	2	2	2	12	57		BLN,BLT			[-]
	48 Hours	2	3	30	1	5	2	2	2	12	47		BLN,BLT			
	72 Hours	3	3	45	1	5	3	2	2	14	64		BLN,BLT,SCE			
	7 Days	3	3	45	1	5	2	2	2	12	62		BLN,VAS-2			
	10 Days	3	2	30	1	5	2	2	2	12	47		BLN,VAS-2			
	14 Days	2	1	10	0	0	1	2	0	6	16		BLN,VAS-2			
	21 Days	2	1	10	0	0	1	1	0	4	14		VAS-1			
56193/F 2.502	1 Hour	3	2	30	1	5	2	1	2	10	45		BLN,BLT,SCE			
	24 Hours	2	4	40	1	5	2	2	2	12	57		BLN,BLT			[-]
	48 Hours	2	4	40	1	5	3	2	2	14	59		BLN,BLT,SCE			
	72 Hours	3	4	60	1	5	3	2	2	14	79		BLN,BLT,VAS-1			
	7 Days	3	4	60	1	5	2	2	2	12	77		BLN,BLT,VAS-3			
	10 Days	3	4	60	1	5	2	2	2	12	77		BLN,BLT,VAS-3			
	14 Days	4	2	40	1	5	2	2	1	10	55		BLN,BLT,VAS-3,RAC,APM			
	21 Days	4	2	40	0	0	1	1	0	4	44		BLN,VAS-2 RAC,APM			

*See Protocol Appendix A for definition of codes; BLN = Blanching on cornea; BLT = Blanching on lower lid of conjunctival tissue; APM = Apparent pseudopterygium.

SLI STUDY NO.: 3255.142
CLIENT: ELF ATOCHEM

TABLE 1
A PRIMARY EYE IRRITATION STUDY IN RABBITS
INDIVIDUAL OCULAR IRRITATION SCORES

PAGE 2

Animal No./Sex Body Weight (kg)	Scoring Interval	Cornea			Iris			Conjunctivae			Test Eye*		Control Eye*	
		O A OxAx5			I lx5			R S D (R+S+D)/2			Fluorescein Exam		Fluorescein Exam	
											Secondary Ocular Findings		Secondary Ocular Findings	
56194/F 2.740	1 Hour	3	3	45	1	5	2	2	2	12				
	24 Hours	2	3	30	1	5	3	2	2	14				
	48 Hours	2	3	30	1	5	3	2	1	12				
	72 Hours	2	3	30	1	5	3	2	1	12				
	7 Days	0	0	0	0	0	2	2	0	8				
	10 Days	0	0	0	0	0	1	1	0	4				
	14 Days	0	0	0	0	0	1	1	0	4				
	21 Days	0	0	0	0	0	1	0	0	2				
56196/F 2.565	1 Hour	3	2	30	1	5	2	2	2	12				
	24 Hours	2	4	40	1	5	2	2	2	12				
	48 Hours	2	4	40	1	5	3	2	2	14				
	72 Hours	3	3	45	1	5	3	3	1	14				
	7 Days	2	1	10	0	0	2	2	0	8				
	10 Days	2	1	10	0	0	2	2	0	8				
	14 Days	2	1	10	0	0	1	1	0	4				
	21 Days	1	1	5	0	0	1	1	0	4				
											Total			
											62			
											49			
											47			
											47			
											8			
											4			
											4			
											2			
											47			
											57			
											59			
											64			
											18			
											18			
											14			
											9			

*See Protocol Appendix A for definition of codes; BLN = Blanching on nictitating membrane; BLT = Blanching on lower lid of conjunctival tissue.

TABLE 1
A PRIMARY EYE IRRITATION STUDY IN RABBITS
INDIVIDUAL OCULAR IRRITATION SCORES

Animal No./Sex Body Weight (kg)	Scoring Interval	Cornea		Iris		Conjunctivae			Test Eye*			Control Eye*		
		O A		Ox	Ax5	I	Ix5	R	S	D (R+S+D)/2	Fluorescein Exam	Secondary Ocular Findings	Fluorescein Exam	Secondary Ocular Findings
		Total		Total		Total		Total		Total		Total		
56162/M	1 Hour	3	3	45	1	5	2	2	2	12	62	BLN,BLT,SCE		
2.705	24 Hours	2	3	30	1	5	2	2	2	12	47	BLN,BLT	[-]	
	48 Hours	3	3	45	1	5	3	2	2	14	64	BLN,BLT,SCE		
	72 Hours	3	3	45	1	5	3	2	2	14	64	BLN,BLT		
	7 Days	2	1	10	0	0	2	2	0	8	18	BLN,BLT,VAS-2		
	10 Days	2	1	10	0	0	2	2	0	8	18	BLN,BLT,VAS-1		
	14 Days	2	1	10	0	0	1	1	0	4	14	VAS-1		
	21 Days	1	1	5	0	0	1	0	0	2	7	[-]	VAS-1	

*See Protocol Appendix A for definition of codes; BLN = Blanching on nictitating membrane; BLT = Blanching on lower lid of conjunctival tissue.

Mean Ocular Scores

1 Hour	-	53.83
24 Hours	-	52.67
48 Hours	-	54.17
72 Hours	-	61.17
7 Days	-	43.67
10 Days	-	33.67
14 Days	-	19.83
21 Days	-	13.83

SLI Study No. 3255.142

(18)

APPENDIX A

Protocol

**A PRIMARY EYE IRRITATION STUDY IN RABBITS
WITH LITHIUM TETRAFLUOROBORATE**

PROTOCOL

Springborn Study No. 3255.142

Protocol No.: ELFATO/EI-1
Issue Date: February 1997

Springborn Laboratories, Inc. (SLI)
Health and Environmental Sciences
640 North Elizabeth Street
Spencerville, Ohio 45887

Deborah A. Douds, M.S.
Study Director

For

Elf Atochem North America, Inc.
2000 Market Street
Philadelphia, PA 19103

SLI Study No. 3255.142

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I. PURPOSE

The purpose of the study is to assess the irritant and/or corrosive effects of a test article in rabbits when administered by a single ocular dose. This study is intended to provide information on the potential health hazards of the test article with respect to ocular exposure. Data from this study may serve as a basis for classification and/or labeling of the test article.

II. RESPONSIBILITIES

A. Sponsor

Elf Atochem North America, Inc.
2000 Market Street
Philadelphia, PA 19103

B. Sponsor's Representative

Roy Bannister, Ph.D.
Phone: (215) 419-5875
FAX: (215) 419-5800

C. Testing Location

Springborn Laboratories, Inc.
Health and Environmental Sciences
553 North Broadway
Spencerville, OH 45887
Phone: (419) 647-4196
FAX: (419) 647-6560

D. Personnel Responsibilities

1. Deborah A. Douds, M.S.
Study Director/Assistant Manager of Acute Toxicology
2. Kimberly L. Bonnette, M.S., LATG
Alternate Contact/Manager of Acute Toxicology
3. Todd N. Merriman, B.S., LATG
Toxicologist
4. Robert C. Springborn, Ph.D.
Chairman, President and CEO
5. Malcolm Blair, Ph.D.
Vice President and Managing Director
6. Rusty E. Rush, M.S., LAT, DABT
Associate Director of Toxicology
7. J. Dale Thurman, D.V.M., M.S., DACVP
Director of Pathology
8. Anita M. Bosau
Director of Compliance Assurance

III. PROPOSED STUDY SCHEDULE

- | | |
|---------------------------------|--|
| A. Initiation of In-Life Phase: | September 1997 |
| B. Completion of In-Life Phase: | September 1997 |
| C. Audited Report Date: | 4 Weeks Following Sponsor's Approval for Study Termination |

IV. TEST ARTICLE IDENTIFICATION

A. Test Article

1. Sponsor's Identification

Lithium Tetrafluoroborate

2. SLI Test Article Identification Number

S97.015.3255

3. Characteristics

The Sponsor is responsible for any necessary evaluations related to identity, strength, purity, composition, stability and method of synthesis of the test material according to 40 CFR 792.105 and OECD [Annex 2 C(81)30]. Any special storage conditions for the test article will be supplied by the Sponsor.

4. Handling Precautions

Safety data regarding the test article should be provided by the Sponsor [Material Safety Data Sheet (MSDS), if available]. Technical personnel are required to read this information prior to handling the test article. Any question concerning this information should be referred to the Study Director.

Additional safety and handling information may be provided by the Study Director and/or Sponsor. Minimum safety requirements include safety glasses, impervious gloves, and laboratory wear. An MSDS shall also be available for any other chemical entities utilized in the conduct of this study.

B. Retention Sample

A 1 g or 1 mL retention sample of each lot of the test article will be taken for studies with an experimental period of 4 weeks or longer. Otherwise, the Sponsor will be responsible for maintaining a retention sample of the test article.

C. Test Article Disposition

The test article will be returned to the Sponsor following completion of all studies with the test article(s) unless otherwise instructed by the Sponsor.

D. Method of Test Article Preparation

Liquids, gels and pastes are generally administered as received from the Sponsor. Solids and powders are generally ground and sieved prior to test use. This may be accomplished by grinding the material in a mortar and pestle and passing the material through a No. 40 mesh sieve. The weight of processed test article that occupies a volume of 0.1 mL will be determined by measuring a convenient volume (at least 2 mL) of the powder in a suitable volumetric container. The powder will be gently compacted by tapping the measuring container. The test article dose per eye will then be calculated (weight equivalent of 0.1 mL, not to exceed 0.1 g). The test article will be prepared and/or dispensed fresh on the day of dosing. The method of preparation will be documented in the raw data and presented in the final report.

V. TEST SYSTEM

A. Justification of the Test System

1. The rabbit is the preferred species for primary eye irritation testing by various U.S. and International regulatory agencies.
2. The New Zealand White rabbit has been shown to be sensitive to the irritant/corrosive effects of a variety of drugs and chemicals. Therefore, this species and strain is a reasonable alternative to larger mammals for primary eye irritation testing of drugs and chemicals for human safety assessment.
3. The New Zealand White rabbit has been used extensively for eye irritation testing. Thus, data from this study may be compared and contrasted to other studies performed in New Zealand White rabbits.
4. Historical information concerning New Zealand White rabbits is available at SLI and in the published literature.

5. Healthy, outbred New Zealand White rabbits may be obtained from reliable, USDA approved and regulated suppliers.
6. The laboratory rabbit may be safely handled and manipulated by trained technical personnel.

SLI has conducted literature searches through Medline, Toxline and Bioethics and there are no generally accepted validated alternatives to this test. In a 1989 position paper prepared by the Animals in Research Committee of The Society of Toxicology (SOT) and approved by the SOT Council, it was concluded that "none of these proposed models are yet validated or evaluated for a broad range of chemical moieties, and none can be relied upon to provide the scientific reliability or predictive accuracy which would be required for a new test for regulatory of legal acceptability" [1].

B. Justification of the Route of Exposure and Number of Animals

1. Ocular administration of the test substance was selected since this is a potential route of human exposure.
2. Since New Zealand White rabbits have no pigment and have an easily accessible ocular area, substances may be accurately instilled and any resulting effects easily observed.
3. The number of animals used on this study will be consistent with the guidelines published by a number of U.S. and International regulatory agencies including EPA-FIFRA, EPA-TSCA, EEC, OECD and MAFF.

C. Description

1. Species

Rabbit

2. Strain

New Zealand White

3. Source

Myrtle's Rabbitry or another USDA approved supplier

4. Age and Body Weight Range

Adult, approximately 2.0 to 3.5 kg (prior to dosing on day 0).

5. Number of Animals/Sex on Study

6 rabbit test (males and/or females)

D. Method of Identification

Plastic ear tags displaying unique identification numbers will be used to individually identify the animals. The cage cards will display at least the study number, animal number, and sex and will be affixed to each cage.

E. Animal Husbandry**1. Housing**

The animals will be housed individually in suspended stainless steel cages. All housing and care will be based on the standards recommended by the Guide for the Care and Use of Laboratory Animals [2].

2. Environment

The environmental conditions for the animal room will be set to maintain room temperature and relative humidity ranges of $67 \pm 6^{\circ}\text{F}$ and $50 \pm 20\%$, respectively. Environmental control equipment will be monitored and adjusted as necessary to minimize fluctuations in the animal room environment. Light timers will be set to maintain a 12-hour light/12-hour dark cycle and the room ventilation will be set to produce 10-15 air changes/hour. The room temperature and relative humidity will be recorded a minimum of once daily.

3. Food

PMI Certified Rabbit Chow #5322 (Purina Mills, Inc.) will be provided ad libitum to the animals throughout the study. The lot number and expiration date of each batch of diet used during the study will be recorded. The feed is analyzed and certified by the supplier for nutritional components and environmental contaminants. Dietary limitations for various environmental contaminants, including heavy metals, pesticides, polychlorinated biphenyls and total aflatoxin are set by the manufacturer. Within these limits, there are no contaminants reasonably expected in the diet which would interfere with the conduct of the study. Results of the dietary analyses (Certificates of Analysis) are provided by the manufacturer for each lot of diet. These will be maintained in the laboratory records. Feed that is outside the ranges set for the above mentioned criteria will not be utilized by the testing facility.

4. Water

Municipal tap water following treatment by reverse osmosis will be available ad libitum throughout the study. The purified water will be supplied by an automatic watering system. Monitoring of the drinking water for contaminants will be conducted annually by the testing laboratory and the records will be available for inspection. Levels of contaminants which may be present are not expected to compromise the purpose of the study. The water meets the standards specified under the EPA National Drinking Water Regulations (40 CFR, Part 141).

F. Acclimation

Upon receipt, the animals will be removed randomly from the shipping cartons, examined by qualified personnel, identified with plastic ear tags, and then acclimated to the laboratory conditions for a minimum of 5 days. The animals will be observed daily for overt physical or behavioral abnormalities, general health/moribundity and mortality.

G. Animal Selection

The animals chosen for study use will be arbitrarily selected from healthy stock animals to avoid potential bias. All animals will receive a detailed pretest observation prior to dosing. Only healthy animals will be chosen for study use. Females will be nulliparous and nonpregnant.

VI. EXPERIMENTAL DESIGN AND PROCEDURES

A. Study Group Design

A six rabbit test will be performed. Materials which are determined by the Sponsor to be strong acids ($\text{pH} \leq 2.0$), strong alkalis ($\text{pH} \geq 11.5$) or a material which produces severe dermal irritation need not be tested in a full number of animals due to their predictive corrosive properties. However, at the request of the Sponsor, these materials will be administered to one animal. If no corrosive response is seen during the first 72 hours, the material will be tested on the remaining five animals.

B. Preliminary Examination

On day 0 prior to dosing, both eyes of each animal provisionally selected for test use will be examined macroscopically for ocular irritation with the aid of an auxiliary light source. In addition, the corneal surface will be examined using fluorescein sodium dye. One drop of a fluorescein/physiological saline mixture will be gently dropped onto the superior sclera of each eye. Following an approximate 15 second exposure, the eyes will be thoroughly rinsed with physiological saline. The corneal surface will then be examined for dye retention under a long-wave UV light source. Animals exhibiting ocular irritation, preexisting corneal injury or fluorescein dye retention will not be used on study. All animals found to be acceptable for test use will be returned to their cages until dosing.

C. Dosing

A minimum of one hour after preliminary ocular examination, the test article will be instilled into the conjunctival sac of the right eye of each animal after gently pulling the lower lid away from the eye. Liquids, gels and pastes will be administered at a volume of 0.1 mL. Solids and powders will be administered at a weight equivalent to 0.1 mL volume, not to exceed 0.1 g. Following instillation, the eyelids will be gently held together for approximately one second in order to limit test article loss and the animal returned to its cage. The contralateral eye will remain untreated to serve as a control. Following dosing, the Study Director will be notified by the technician if severe local reactions occur or if the animals exhibit overt clinical indications of pain/distress immediately post-dose. If such is noted, the Sponsor will be contacted to see if the animals should be humanely euthanized.

D. Body Weights

Individual body weights will be obtained for each animal prior to dosing on day 0.

E. Ocular Observations

The eyes will be macroscopically examined with the aid of an auxiliary light source for signs of irritation at 1, 24, 48 and 72 hours after dosing according to the Ocular Grading System presented in Protocol Appendix A which is based on Draize [3]. At the discretion of the Study Director, a biomicroscopic slit-lamp may be utilized to further examine and clarify ocular lesions. Following macroscopic observations at the 24 hour scoring interval, the fluorescein examination procedure will be repeated on all test and control eyes and any residual test article should be gently rinsed from the eye at this time (if possible) using physiological saline. If any fluorescein findings are noted at 24 hours, a fluorescein exam will be conducted on the affected eyes at each subsequent interval until a negative response is obtained and/or until all corneal opacity has cleared, or as directed by the Study Director. If there is no evidence of treatment-related ocular irritation at the 72 hour scoring interval, the study will be terminated. If ocular irritation persists in any test eye, the observation period may be extended for the affected animals (scored on days 7, 10, 14 and 21). Animals requiring an extended observation period will remain on test until the irritation has resolved, permanent injury is evident or the Study Director/Sponsor determines that additional scoring intervals are unnecessary.

F. Clinical Observations

Any unusual observations and mortality will be recorded. General health/mortality checks will be performed twice daily (in the morning and in the afternoon).

G. Unscheduled Deaths and Euthanasia

Any animals dying or euthanized for cause during the study period will be necropsied. The animals will be euthanized by an intravenous injection of sodium pentobarbital. Body cavities (cranial, thoracic, abdominal and pelvic) will be opened and examined. No tissues will be retained.

H. Scheduled Euthanasia

Each surviving animal will be euthanized by an intravenous injection of sodium pentobarbital following its final observation interval. A gross necropsy examination will not be required for surviving animals.

VII. PROTOCOL AMENDMENT

Alterations to this protocol may be made as the study progresses. No changes in the protocol will be made without the specific consent of the Sponsor's Representative. A protocol amendment will be prepared and signed by the Study Director, SLI Quality Assurance and Sponsor's Representative for any such changes.

VIII. DATA REPORTING

One unbound copy of the draft report and two copies of the final report (one bound and one unbound) will be submitted to the Sponsor. The final report will include all information necessary to provide a complete and accurate description and evaluation of the experimental procedures and results.

The report will include at least the following information and data:

- Table of Contents
- Regulatory Compliance
- Summary
- Introduction
- Experimental Design and Test Procedures
- Presentation and Discussion of Results
- Conclusion
- References
- Data Tables
- Protocol and Amendments
- SLI Personnel Responsibilities

IX. ANALYSIS OF DATA

For each group, the ocular irritation score for each parameter (i.e., corneal opacity x area, iritis and conjunctival redness + swelling + discharge) will be multiplied by the appropriate factor (i.e., corneal injury x 5, iritis x 5, conjunctivitis x 2) and the totals added for each animal/interval. The group mean irritation score will then be calculated for each scoring interval based on the number of animals initially dosed in each group. If an animal dies during the study, the total animals in that group will be reduced (by the number of animals dead) for each subsequent scoring interval for the purpose of calculating the mean ocular irritation score for each interval.

The data will be analyzed and summarized in the report based on the definitions presented below:

1. Non Irritant - Following instillation of the test article, none of the test eyes show a positive effect as defined in the Ocular Grading System in Protocol Appendix A.
2. Irritant - Following instillation of the test article, one or more test eyes exhibit a positive effect, but the changes are reversible.
3. Corrosive - One or more test eyes exhibit irreversible changes (i.e., necrosis or ulceration) following instillation of the test article.

X. MAINTENANCE OF RAW DATA, RECORDS AND SPECIMENS

All original data, magnetically encoded records, specimens and reports from this study are the property of the Sponsor. These materials shall be available at SLI to facilitate auditing of the study during its progress and prior to acceptance of the final report. All original paper data, the final report, magnetically encoded records, and any specimens will be transferred to the SLI archives for a period of 10 years. The Sponsor will be contacted prior to the final disposition of these items.

XI. REGULATORY COMPLIANCE

This study may be submitted to and will be conducted in accordance with the EPA-TSCA [4], OECD [5] and EEC [6] guidelines and the principles of the Good Laboratory Practice Standards by the EPA (40 CFR Part 792) and the OECD [Annex 2 C(81)30].

XII. QUALITY ASSURANCE

The study will be inspected at least once during the in-life phase by the Springborn Laboratories, Inc., Quality Assurance Unit to assure compliance with Good Laboratory Practice Standards, SLI's Standard Operating Procedures and for conformance with the protocol and protocol amendments. The final report will be audited prior to submission to the Sponsor to ensure that it completely and accurately describes the test procedures and results of the study.

XIII. USDA ANIMAL WELFARE COMPLIANCE STATEMENT

This study will comply with all applicable sections of the Final Rules of the Animal Welfare Act regulations (9 CFR) and the Public Health Service Policy on Humane Care and Use of Laboratory Animals (OPRR, NIH, 1986). Wherever possible, procedures used in this study have been designed to avoid or minimize discomfort, distress and pain to animals. All methods are described in this study protocol or in written laboratory standard operating procedures. These procedures are based on the most currently available technologies concerning proper laboratory animal use and management. Methods of euthanasia used during this study are in conformance with the above referenced regulations and the American Veterinary Medical Association Panel on Euthanasia [7]. This protocol has been reviewed and approved by Springborn Laboratories, Inc. Institutional Animal Care and Use Committee (IACUC) for a maximum of 18 animals. Prior IACUC approval will be obtained for repeated studies.

This study is being conducted to evaluate potential irritant effects of the test article and potential reversibility of such effects. Following dosing, the Study Director will be notified by the technician if severe local reactions occur or if the animals exhibit overt clinical indications of pain/distress post-dose or if delayed severe ocular or clinical changes subsequently develop. If severe reactions are noted, the Study Director will contact the Facility Veterinarian and Sponsor to consider an appropriate course of action. In the event that the Sponsor cannot be contacted, the Study Director and Facility Veterinarian may authorize treatment or euthanasia of the animals. In general, the ocular tissue will not be anesthetized prior to or following dosing since these substances have been shown to inhibit the blink and/or tear response which may alter the irritation response. Furthermore, anesthetic agents may interact with and/or dilute the test article and thereby alter the experimental results. However, if it is suspected that the test article may induce more than transient pain/distress based on existing information, preanesthesia will be considered. In such circumstances, the Study Director and/or Facility Veterinarian will consult with the Sponsor to devise an appropriate study plan.

XIV. DECLARATION OF INTENT

This study will be listed on the SLI Quality Assurance Master Schedule for the EPA.

XV. GENERIC PROTOCOL APPROVAL

The Sponsor's signature below documents that there are no acceptable non-animal alternatives for this study, and that since this study is required by the relevant supervising government agency, it does not unnecessarily duplicate any previous experiments.

Kimberly L. Bonnette
Kimberly L. Bonnette, M.S., LATG
Manager of Acute Toxicology (SLI)

Date: 3/4/97

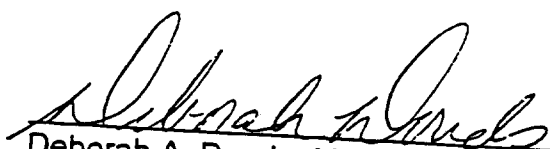
Christopher W. Wilson
Christopher W. Wilson, B.S.
Supervisor of Quality Assurance (SLI)

Date: 3/5/97

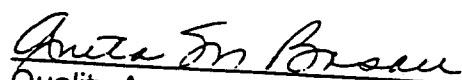
Roy B. Bangerter
Roy Bangerter, Ph.D.
Sponsor's Representative
(Principal Investigator)

Date: 2/28/97

XVI. STUDY SPECIFIC PROTOCOL APPROVAL


Deborah A. Douds, M.S.
Study Director (SLI)

Date: 9/4/97


Anita En Brosau
Quality Assurance Unit (SLI)

Date: 9/4/97

XVI. REFERENCES

1. SOT Position Paper, "Comments on the LD50 and Acute Eye and Skin Irritation Tests, Fundamental and Applied Toxicology 13, 621-623, 1993.
2. Guide for the Care and Use of Laboratory Animals, DHHS Publication No. (NIH) 96-03, 1996.
3. Draize, J.H., Appraisal of the Safety of Chemicals in Foods, Drugs and Cosmetics, The Association of Food and Drug Officials of the United States, 49-51, 1959.
4. Toxic Substances Control Act Test Guidelines, 40 CFR Part 798, Subpart E, Section 798.4500, July 1, 1992.
5. OECD Guidelines for the Testing of Chemicals, Section 4, Health Effects, Subsection 405, February 24, 1987.
6. The EEC Guidelines Part B: Method for the Determination of Toxicity, No. L 383 A/127, B.5, December 29, 1992.
7. 1993 Report of the American Veterinary Medical Assoc. Panel on Euthanasia, JAVMA, Vol. 202, No. 2, pp. 229-249, January 15, 1993.

PROTOCOL APPENDIX A

Ocular Grading System

(O) CORNEAL OPACITY-DEGREE OF DENSITY (AREA MOST DENSE TAKEN FOR READING)	
OBSERVATION	CODE
No ulceration or opacity	0
Scattered or diffuse areas of opacity (other than slight dulling of normal luster), details of iris clearly visible	1*
Easily discernible translucent area, details of iris slightly obscured	2*
Nacreous (opalescent) area, no details of iris visible, size of pupil barely discernible	3*
Opaque cornea, iris not discernible through opacity	4*

(A) AREA OF CORNEA INVOLVED (TOTAL AREA EXHIBITING ANY OPACITY, REGARDLESS OF DEGREE)	
OBSERVATION	CODE
No ulceration or opacity	0
One quarter (or less) but not zero	1
Greater than one quarter, but less than half	2
Greater than half, but less than three quarters	3
Greater than three quarters, up to whole area	4

Cornea Score = O x A x 5

Total Maximum = 80

(I) IRITIS	
OBSERVATION	CODE
Normal	0
Markedly deepened rugae (folds above normal), congestion, swelling, moderate circumcorneal hyperemia or injection, any or all of these or combination of any thereof, iris is still reacting to light (sluggish reaction is positive)	1*
No reaction to light, hemorrhage, gross destruction (any or all of these)	2*

Iris Score = I x 5

Total Maximum = 10

*Positive Effect.

PROTOCOL APPENDIX A—(Continued)

Ocular Grading System

(R) CONJUNCTIVAL REDNESS (REFERS TO PALPEBRAL AND BULBAR CONJUNCTIVAE EXCLUDING CORNEA AND IRIS)	
OBSERVATION	CODE
Blood vessels normal	0
Some blood vessels definitely hyperemic (injected) above normal (slight erythema)	1
Diffuse, crimson color, individual vessels not easily discernible (moderate erythema)	2*
Diffuse beefy red (marked erythema)	3*

(S) CONJUNCTIVAL SWELLING (LIDS AND/OR NICTATING MEMBRANE)	
OBSERVATION	CODE
No swelling	0
Any swelling above normal (includes nictitating membrane, slightly swollen)	1
Obvious swelling with partial eversion of lids	2*
Swelling with lids about half closed	3*
Swelling with lids more than half closed	4*

(D) CONJUNCTIVAL DISCHARGE	
OBSERVATION	CODE
No discharge	0
Any amount different from normal (does not include small amounts observed in inner canthus of normal animals)	1
Discharge with moistening of the lids and hairs just adjacent to lids	2
Discharge with moistening of the lids and hairs and considerable area around the eye	3

Conjunctival Score = (R + S + D) x 2

Total Maximum = 20

*Positive Effect.

PROTOCOL APPENDIX A--(Continued)

Ocular Grading System

CORNEAL NEOVASCULARIZATION		
OBSERVATION	CODE	DEFINITION
Neovascularization - Very Slight	VAS-1	Total area of vascularized corneal tissue is < 10% of corneal surface
Neovascularization - Mild	VAS-2	Total area of vascularized corneal tissue is > 10% but < 25% of corneal surface
Neovascularization - Moderate	VAS-3	Total area of vascularized corneal tissue is > 25% but < 50% of corneal surface
Neovascularization - Severe	VAS-4	Total area of vascularized corneal tissue is > 50% of corneal surface

SECONDARY OCULAR FINDINGS		
OBSERVATION	CODE	DEFINITION
Sloughing of the corneal epithelium	SCE	Corneal epithelial tissue is observed to be peeling off the corneal surface.
Corneal bulging	CB	The entire corneal surface appears to be protruding outward further than normal.
Slight dulling of normal luster of the cornea	SDL	The normal shiny surface of the cornea has a slightly dulled appearance.
Raised area on the corneal surface	RAC	A defined area on the corneal surface that is raised above the rest of the cornea. This area is generally associated with neovascularization and has a off-white to yellow color.
Corneal edema	CE	The cornea has a swollen appearance.
Test article present in eye	TAE	Apparent residual test article is observed on the eye or in the conjunctival sac/inner canthus.
Observation confirmed by slit lamp	OCS	A slit lamp examination was performed to confirm the initial observation.
Corneal mineralization	CM	Small white or off-white crystals that are observed in the corneal tissue.

PROTOCOL APPENDIX A--(Continued)

Ocular Grading System

FLUORESCEIN EXAMINATION OF CORNEA	
OBSERVATION	CODE
<u>Fluorescein Dye Retention</u> Fluorescein dye retention associated with the area of corneal opacity Fluorescein dye retention is not associated with any other finding	FAO FNF
<u>Negative Results</u> No fluorescein retention is observed	(-)
<u>Secondary Ocular Findings</u> Superficial mechanical abrasion to the cornea observed during the fluorescein examination period Fine stippling on the cornea observed during the fluorescein examination procedure	MI ST

POST-DOSE CLINICAL OBSERVATIONS	
OBSERVATION	CODE
Animal vocalized following dosing	VOC
Animal excessively pawed test eye following dosing	PAW
Animal exhibited excessive hyperactivity following dosing	HYP
Animal exhibited excessive head tilt following dosing	HT
Animal exhibited excessive squinting of test eye following dosing	SQ

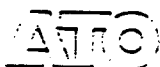
Any additional findings will be noted in the raw data and in the final report.

SLI Study No. 3255.142

(40)

APPENDIX B

Certificate of Analysis
(Provided by the Sponsor)

elf atochem

(41)
OZARK-MAHONING COMPANY
AN ELF ATOCHEM NORTH AMERICA, INC. SUBSIDIARY

AUG 13 1997

CERTIFICATE OF ANALYSIS5101 WEST 21ST STREET
TULSA, OKLAHOMA 74107PHONE: (918) 583-0851
TELEFAX: (918) 583-0858

SHIPMENT OF LITHIUM TETRAFLUOROBORATE TO:

ORDER NUMBER:

LOT NUMBER:

AQ-2-166(F)

AMOUNT:

ANALYSIS:

% of Theory

> 99.5

% F Found

80.87


% Free F Found

0.057

ANALYTICAL REPORT NUMBER:

RC112

We certify that this material meets all specifications of the Ozark-Mahoning Company.


Analytical Department
August 4, 1997

TECHNICAL INFORMATION AND DATA REGARDING THE COMPOSITION, PROPERTIES OR USE OF THE PRODUCTS DESCRIBED HEREIN IS BELIEVED RELIABLE, HOWEVER NO REPRESENTATION OR WARRANTY IS MADE WITH RESPECT THERETO EXCEPT AS MADE BY ELF ATOCHEM IN WRITING AT TIME OF SALE. ELF ATOCHEM CANNOT ASSUME RESPONSIBILITY FOR ANY PATENT LIABILITY WHICH MAY ARISE FROM THE USE OF ANY PRODUCT IN A PROCESS, MANNER OR FORMULA NOT DESIGNED BY ELF ATOCHEM.

APPENDIX C

Individual Clinical Observations

SLI STUDY NO.: 3255.142
CLIENT: ELF ATOCHEM

A PRIMARY EYE IRRITATION STUDY IN RABBITS
INDIVIDUAL CLINICAL OBSERVATIONS
(POSITIVE FINDINGS)

PAGE 1

Group	Animal No./Sex	Clinical Observations
No-Rinse	55983/M	Animal vocalized following dosing: Day 0 Hairloss - around right eye - moderate: Days 7, 10, 14
	56192/F	Animal vocalized following dosing: Day 0 Few feces: Day 2 Low food consumption: Day 2 Hairloss - around right eye - slight: Day 14
	56193/F	Animal vocalized following dosing: Day 0 Hairloss - around right eye - moderate: Days 7, 10, 14 Hairloss - around right eye - slight: Day 21
	56194/F	Animal vocalized following dosing: Day 0 Hairloss - around right eye - slight: Days 2, 3, 7, 10
	56162/M	Animal vocalized following dosing: Day 0 Hairloss - around right eye - slight: Day 14

APPENDIX D

SLI Personnel Responsibilities

SLI PERSONNEL RESPONSIBILITIES

Deborah A. Douds, M.S.	Study Director/Assistant Manager of Acute Toxicology
Kimberly L. Bonnette, M.S., LATG	Alternate Contact/Manager of Acute Toxicology
Malcolm Blair, Ph.D.	Vice President and Managing Director
Joseph C. Siglin, Ph.D., DABT	Director of Toxicology
Pamela S. Smith, ALAT	Primary Technician/Unit Leader
Delores P. Knippen	Supervisor of Pharmacy
Jan K. Severt, B.S., ALAT	Supervisor of Acute Report Preparation
Anita M. Bosau	Director of Compliance Assurance
Christopher W. Wilson, B.S.	Supervisor of Acute Quality Assurance
Deanna M. Talerico	Supervisor of Nonacute Quality Assurance
J. Dale Thurman, D.V.M., M.S., DACVP	Director of Pathology